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Production of alcohol-based quorum sensing molecules and their role in biofilm formation and sliding motility of the dairy-important yeast *Debaryomyces hansenii*

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Aim
The aim of the present study was to examine aromatic alcohols including phenylethanol, tyrosol and tryptophol and the sesquiterpene farnesol for their potential quorum sensing effects in the dairy-important yeast *Debaryomyces hansenii*

Introduction
Many yeasts are dimorphic, which means that they are able to undergo a transition from a unicellular yeast form to a filamentous form or vice-versa. The yeast-to-mycelium transition can be induced by environmental factors e.g. temperatures below or over the optimal range, limited availability of oxygen, high osmolality, pH or substrate limitation including nitrogen starvation. In addition, quorum sensing (the process by which microorganisms communicate by signalling molecules in a cell density dependent manner) has reported to be involved in the morphogenesis of yeasts. The sesquiterpene farnesol has been found to be a signalling molecule inhibiting both the yeast-to-hyphal shift and biofilm formation of *Candida albicans*. In addition, the aromatic alcohol tyrosol has been shown due to its promotion of hyphal development to shorten lag phase time of *C. albicans*. Furthermore, the aromatic alcohols phenylethanol and tryptophol have been found to be signalling molecules in *Saccharomyces cerevisiae*, where they stimulate pseudohyphal growth. Furthermore, phenylethanol was found to stimulate invasive growth. Addition of tryptophol resulted in an even greater invasive growth, whereas tryptophol alone did not have an effect on invasive growth. *C. albicans* does also produce phenylethanol and tryptophol, but if a function, it is different, from what is observed in *S. cerevisiae*.

Material and methods
Three strains of *Debaryomyces hansenii* (type strain CBS767 and the dairy isolates D18335 and MD02) were included. *Candida albicans* (CBS8758) and *Saccharomyces cerevisiae* (CBS1171) were included as reference strains. At standard conditions, yeast cultures were grown in yeast nitrogen base (YNB) without amino acids and ammonium sulfate supplemented with 2% glucose and 5 mM L-Proline (pH 4.3) at 25°C with shaking at 120 rpm. At appropriate time intervals, cell-free supernatants were prepared and analysed by their content of phenylethanol, tyrosol and tryptophol by ESI⁺ with the triple quadrupole operating in multiple reaction monitoring mode with two transitions per compound. Quantification was done using ²H₅-1-phenylethanol as internal standard (10 ng/mL) and using peak areas as response factor relative to ²H₅-1-phenylethanol. Farnesol was determined in a similar fashion applying farnesol as internal standard.

Biofilm formation was measured by the crystal violet method. Sliding motility was measured by inoculation of yeast cells in the centre of YPD plates with 0.3% agar, and growth was followed for 14 days. To induce pseudohyphal growth, yeast cells were restreaked on SLAD agar. Plates were incubated at 25°C for 10 days. Colony morphology photographed under a microscope.

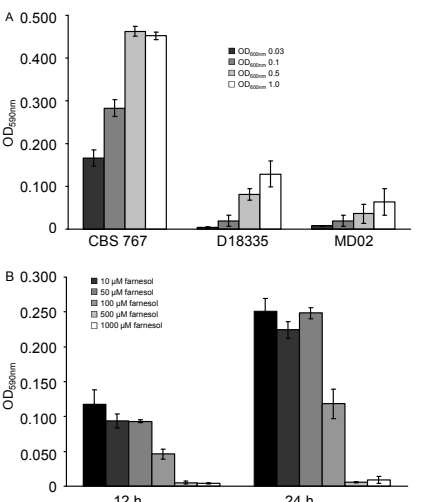


Figure 2. A: Biofilm formation of *D. hansenii* (CBS767, D18335 and MD02) after 24 h of incubation. B: Influence of farnesol on biofilm formation of *D. hansenii* (CBS767).

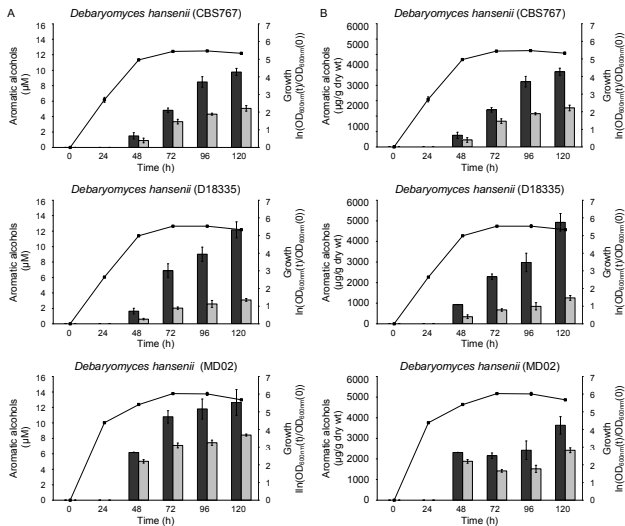


Figure 1. Production of phenylethanol and tyrosol (shown in bars) in *Debaryomyces hansenii* (CBS767, D18335 and MD02) at standard conditions. Growth (ln(OD_{600nm}(t)/OD_{600nm}(0))), t refers to the time.

Table 1. Effects of environmental conditions on phenylethanol and tyrosol production in *Debaryomyces hansenii* (CBS767). Fold inductions/repressions are based on alcohols concentrations in μM determined after 120 h of fermentation.

	Fold inductions/repressions	
	Phenylethanol	Tyrosol
Aromatic amino acid (50 μg/ml)		
Phenylalanine	22 ± 2.3	2.7 ± 0.60
Tyrosine	1.5 ± 0.18	87 ± 15
Tryptophan	2.4 ± 0.75	5.3 ± 0.10
Ammonium sulfate		
50mM	0.78 ± 0.011	nd
100mM	0.82 ± 0.23	nd
500mM	0.77 ± 0.069	0.46 ± 0.00088
1000mM	0.86 ± 0.075	1.7 ± 0.28
5000mM	0.63 ± 0.11	0.30 ± 0.16
37000mM	0.63 ± 0.015	0.32 ± 0.042
NaCl		
2% (w/v)	0.59 ± 0.0035	0.63 ± 0.064
4% (w/v)	0.74 ± 0.067	0.66 ± 0.14
6% (w/v)	0.51 ± 0.015	0.48 ± 0.097
8% (w/v)	0.34 ± 0.047	0.23 ± 0.0051
pH		
5.0	0.80 ± 0.063	0.80 ± 0.0050
6.0	0.87 ± 0.27	1.8 ± 0.15
7.0	nd	0.31 ± 0.11
8.0	nd	nd
9.0	nd	nd
Temperature		
15°C	0.85 ± 0.55	0.81 ± 0.43
Aromatic alcohols (1000 μM)		
Phenylethanol	-	3.8 ± 0.38
Tyrosol	1.2 ± 0.18	-
Tryptophol	1.2 ± 0.20	5.0 ± 0.94
nd not detectable		

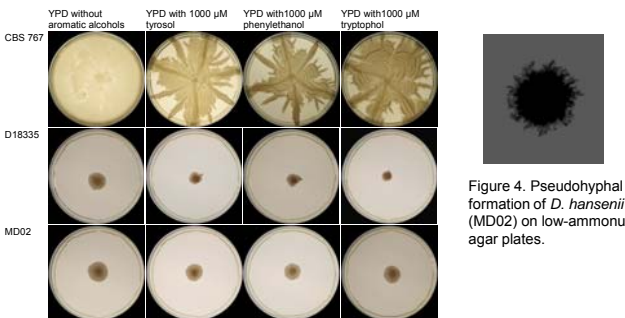


Figure 3. Sliding motility of *D. hansenii* (CBS767, D18335 and MD02) on YPD with 0.3% agar.

Production of alcohols at standard conditions
D. hansenii (CBS 767, D18335 and MD02) were found to produce increasing concentrations of phenylethanol and tyrosol in the μM range during growth at standard conditions (Figure 1A). After 120 h of fermentation, the concentrations of phenylethanol and tyrosol were found to be 9.77-12.6 μM and 3.01-8.42 μM, respectively. Concerning tryptophol, this alcohol was in general not detected at standard conditions. For comparison, the concentrations of phenylethanol, tyrosol and tryptophol were 293, 119 and 11.4 μM for *C. albicans* (CBS 8758) and 31.4, 22.8 and 0.198 μM for *S. cerevisiae* (1171) (results not shown). The fact that alcohols were primarily determined from the end of exponential phase indicates that these alcohols are potential quorum sensing molecules in *D. hansenii*.

When calculated as μg/g dry wt, *D. hansenii* (CBS 767 and D18335) produced increasing concentrations of phenylethanol and tyrosol, whereas *D. hansenii* (MD02) produced constant concentrations of the two alcohols (Figure 1B). The fact that *D. hansenii* (CBS 767 and D18335) produced increasing concentrations of phenylethanol and tyrosol during stationary phase indicates that these alcohols besides being produced as potential quorum sensing molecules also are products used for survival of these two *D. hansenii* strains.

Alcohol production at different environmental conditions
The aromatic alcohol production for the three *D. hansenii* strains varied with different environmental conditions. Table 1 shows the changes in alcohol production at different environmental conditions for *D. hansenii* (CBS767) after 120 h of fermentation. Similar results were obtained for *D. hansenii* (D18335 and MD02) (results not shown). Addition of aromatic amino acids (the precursors of aromatic alcohol production) increased aromatic alcohol production in abundance. Phenylethanol production increased 22-fold by addition of phenylalanine, whereas tyrosol production increased 87-fold by the addition of tyrosine. Addition of tryptophan resulted in tryptophol concentrations between 4.20-7.50 μM. Furthermore, alcohols were not only increased by their own precursors e.g. tryptophan increased tyrosol production by more than 5.0-fold. Production of phenylethanol and tyrosol was generally decreased by increasing concentrations of ammonium, NaCl, pH and temperature values. Contrary to phenylethanol, tyrosol production were found to be autostimulated by addition of phenylethanol and tryptophol, as these alcohols increased tyrosol production by more than 3.8-fold.

Biofilm formation, sliding motility and pseudohyphal growth
Among *D. hansenii* strains investigated, especially *D. hansenii* (CBS767) was capable of biofilm formation (Figure 2a). Phenylethanol, tyrosol and tryptophol did in general not increase biofilm formation of *D. hansenii* (CBS767), whereas farnesol concentrations as low as 500 μM completely inhibited *D. hansenii* (CBS767) in formation of biofilm (Figure 2b). Also concerning sliding motility, *D. hansenii* (CBS767) shown a particular ability as this strain overgrew YPD plates with low agar content (Figure 3). The correlation between biofilm formation and sliding motility suggests an intimate connection of the two phenotypes. Addition of phenylethanol, tyrosol and tryptophol as low as 1000 μM resulted in more filamentous growth pattern *D. hansenii* (CBS767) (Figure 3). Contrary, only *D. hansenii* (MD02) was able of pseudohyphal growth on low-ammonium agar plates (Figure 4).

Conclusions
The present study shows that *D. hansenii* produces alcohol-based quorum sensing molecules previously reported for *C. albicans* and *S. cerevisiae*. However, alcohol yields are highly dependent on growth conditions including the availability of aromatic amino acids, ammonium sulphate, NaCl, pH and temperature. Several physiological traits including biofilm formation and sliding motility of *D. hansenii* seems to be regulated by alcohol-based quorum sensing molecules. Knowledge on quorum sensing properties of dairy-relevant microorganisms including *D. hansenii* may be used in optimisation of the cheese ripening processes.

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